

ATP Chemiluminescence Assay Kit

Catalog No: E-BC-F002

Method: Chemiluminescence immunoassay analyzer,
Multifunctional microplate reader

Specification: 96T (Can detect 80 samples without duplication)

Measuring instrument: Chemiluminescence immunoassay
analyzer, Multifunctional microplate reader

Sensitivity: 0.003 $\mu\text{mol/L}$

Detection range: 0.003-10 $\mu\text{mol/L}$

- ▶ Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

General information

▲ Intended use

This kit can be used to measure ATP content in animal tissue and cell samples.

▲ Background

Adenosine 5' -triphosphate (ATP), an organic compound, is a natural nucleotide present in every cell consisting of purine base (adenine), ribose, and three phosphate groups. The content in tissue cells is generally in dynamic equilibrium, which is of great significance to constitute a stable energy supply environment inside the organism. ATP released from many cells is a physiological or pathophysiological response to mechanical stress, hypoxia, inflammation, and some agonists.

▲ Detection principle

Under the catalyzation of luciferase, ATP react with luciferin and emits fluorescence, and the fluorescence intensity is proportional to the concentration of ATP within a certain range.

▲ Kit components & Storage

| Item | Component | Specification | Storage |
|-----------|------------------------------|-----------------|---------------------------------|
| Reagent 1 | Extracting Solution | 50 mL × 2 vials | -20°C , 3 months |
| Reagent 2 | 100 µmol/L Standard Solution | 1 mL × 1 vial | -20°C , 3 months |
| Reagent 3 | Enzyme Solution | 1 mL × 2 vials | -20°C , 3 months, shading light |
| Reagent 4 | Enzyme Diluent | 10 mL × 1 vial | -20°C , 3 months |
| | Black Microplate | 96 wells | No requirement |
| | Plate Sealer | 2 pieces | |

Note: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other.

▲ Materials prepared by users



Instruments

Vortex mixer, Centrifuge, Water bath , Chemiluminescence immunoassay analyzer or multifunctional microplate reader (with the function of detecting chemiluminescence).



Consumptive material

Tips (10 µL, 200 µL, 1000 µL), EP tubes (1.5 mL, 2 mL)



Reagents

Double distilled water

▲ Safety data

Some of the reagents in the kit contain dangerous substances. It should be avoided to touch the skin and clothing. Wash immediately with plenty of water if touching it carelessly. All the samples and waste material should be treated according to the relevant rules of laboratory's biosafety.

▲ Precautions

Before the experiment, please read the instructions carefully, and wear gloves and work clothes.

▲ The key points of the assay

1. Dilute the samples to the optimal concentration for detection if the ATP content of samples exceed the detection range.
2. The sample size of each batch should be less than 30 (including standard wells).
3. Prevent the formulation of bubbles when the supernatant is transferred into the microplate.

Pre-assay preparation

▲ Reagent preparation

1. Bring reagent 1, reagent 2, reagent 4 to room temperature, and place reagent 3 on ice before detection.
2. Preparation of enzyme working solution:
Mix the reagent 3 and reagent 4 at a ratio of 1:5. Prepare the needed fresh solution before use.

▲ Sample preparation

Tissue sample

Weigh the tissue accurately, cut into pieces, add 9 times the volume of reagent 1 according to the ratio of Weight (g): Volume (mL) =1:9. Mechanical homogenate the sample in ice water bath. Then incubate in boiling water bath for 2 min, cool with the running water and centrifuge at 10000 g for 10 min, then take the supernatant and preserve it on ice for detection.

Cells sample

Collect the cells and add reagent 1 at a ratio of cells number (2×10^6): volume (mL) =1: 0.3. Then incubate in boiling water bath for 10 min, cool the tubes to room temperature with running water. Centrifuge at 10000 g for 10 min at 4°C, then take the supernatant for detection.

▲ Dilution of sample

It is recommended to take 2~3 samples with expected large difference to do pre-experiment before formal experiment and dilute the sample according to the result of the pre-experiment and the detection range (0.003-10 $\mu\text{mol/L}$).

The recommended dilution factor for different samples is as follows (for reference only):

| Sample type | Dilution factor |
|------------------------------------|-----------------|
| 10% Mouse heart tissue homogenate | 1 |
| 10% Mouse kidney tissue homogenate | 1 |
| 10% Mouse muscle tissue homogenate | 1 |
| 10% Mouse liver tissue homogenate | 1 |
| 10% Mouse brain tissue homogenate | 1 |
| 10% Mouse lung tissue homogenate | 1 |

Note: The diluent is reagent 1.

| Assay protocol | |
|---------------------|---------|
| Ambient temperature | 25-30°C |

Instructions for the use of transferpette

- (1) Equilibrate the pipette tip in that reagent before pipetting each reagent.
- (2) Don't add the liquid outside the tips into the reaction system when pipetting each reagent.

Assay protocol

▲ Plate set up

| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
|---|---|---|----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| A | A | A | S1 | S9 | S17 | S25 | S33 | S41 | S49 | S57 | S65 | S73 |
| B | B | B | S2 | S10 | S18 | S26 | S34 | S42 | S50 | S58 | S66 | S74 |
| C | C | C | S3 | S11 | S19 | S27 | S35 | S43 | S51 | S59 | S67 | S75 |
| D | D | D | S4 | S12 | S20 | S28 | S36 | S44 | S52 | S60 | S68 | S76 |
| E | E | E | S5 | S13 | S21 | S29 | S37 | S45 | S53 | S61 | S69 | S77 |
| F | F | F | S6 | S14 | S22 | S30 | S38 | S46 | S54 | S62 | S70 | S78 |
| G | G | G | S7 | S15 | S23 | S31 | S39 | S47 | S55 | S63 | S71 | S79 |
| H | H | H | S8 | S16 | S24 | S32 | S40 | S48 | S56 | S64 | S72 | S80 |

Note: A-H, standard wells; S1-S80, sample wells.

▲ Operating steps

1. The preparation of standard curve
Dilute 100 $\mu\text{mol/L}$ standard solution with reagent 1 to a serial concentration. The recommended dilution gradient is as follows: 0, 0.5, 1, 2, 4, 6, 8, 10 $\mu\text{mol/L}$.
2. The measurement of samples
 - 1) **Standard well:** Add 100 μL of enzyme working solution into the corresponding well and stand for 5 min.
Sample well: Add 100 μL of enzyme working solution into the corresponding well and stand for 5 min.
 - 2) **Standard well:** Add 100 μL of standard with different concentrations into standard well, and mix fully immediately.
Sample well: Add 100 μL of sample supernatant into sample well, and mix fully immediately.
 - 3) Measure the fluorescence values of each well by the chemiluminescence immunoassay analyzer or multifunctional microplate reader.

▲ Operation table

| | Standard well | Sample well |
|---------------------------------------------------------------------------------------------------------------------------------------------------------|---------------|-------------|
| Enzyme working solution (μL) | 100 | 100 |
| Stand for 5 min. | | |
| Standard with different concentrations (μL) | 100 | |
| Supernatant of sample (μL) | | 100 |
| Mix fully immediately. Measure the fluorescence values of each well by the chemiluminescence immunoassay analyzer or multifunctional microplate reader. | | |

▲ Calculation

Plot the standard curve by using fluorescence value (F) of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve with graph software (or EXCEL). The concentration of the sample can be calculated according to the formula based on the F value of sample. The standard curve is: $y = ax + b$.

For tissue sample:

$$\text{ATP content } (\mu\text{mol/kg wet tissue}) = (\Delta F - b) \div a \times f \div m \times V$$

For cells sample:

$$\text{ATP content } (\mu\text{mol}/1 \times 10^6) = (\Delta F - b) \div a \times f \div n \times V$$

Note:

y: $F_{\text{Standard}} - F_{\text{Blank}}$ (F_{Blank} is the fluorescence value when the standard concentration is 0)

x: The concentration of Standard.

a: The slope of standard curve.

b: The intercept of standard curve.

ΔF : The absolute fluorescence value of sample, $F_{\text{Sample}} - F_{\text{Blank}}$.

f: Dilution factor of sample before tested.

m: wet weight of sample, 0.05 g is recommended.

v: the volume of reagent 1 in sample preparation step.

n: the number of cells. For example, the number of cells is 5×10^6 , N is 5.

▲ Notes

1. This kit is for research use only.
2. Instructions should be followed strictly, changes of operation may result in unreliable results.
3. The validity of kit is 3 months.
4. Do not use components from different batches of kit.

Appendix I Performance characteristics

| Appendix I Performance characteristics | | | |
|----------------------------------------|----------------------------|----------------------------|-----|
| Detection range | 0.003-10 $\mu\text{mol/L}$ | Average inter-assay CV (%) | 2.2 |
| Sensitivity | 0.003 $\mu\text{mol/L}$ | Average inter-assay CV (%) | 6.5 |
| Average recovery rate (%) | 102 | | |

▲ Example analysis

For mouse lung tissue, take 0.05 g of fresh mouse lung sample and carry the assay according to the operation table. The results are as follows:

standard curve: $y = 10627x + 770.87$, the average F value of the sample is 9496, the average F value of the blank is 76, and the calculation result is:

$$\begin{aligned}\text{ATP content } (\mu\text{mol/kg wet weight}) &= \frac{(9496 - 76 - 770.87)}{10627} \div 0.05 \times 0.45 \\ &= 7.32 \mu\text{mol/kg wet weight}\end{aligned}$$